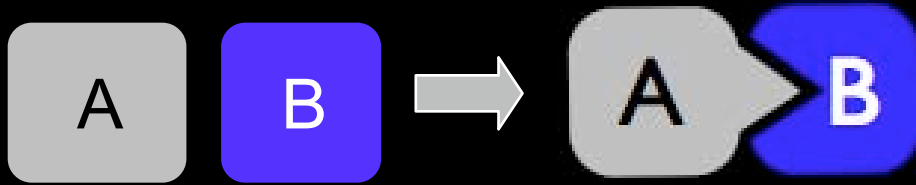


Protein Interface Design

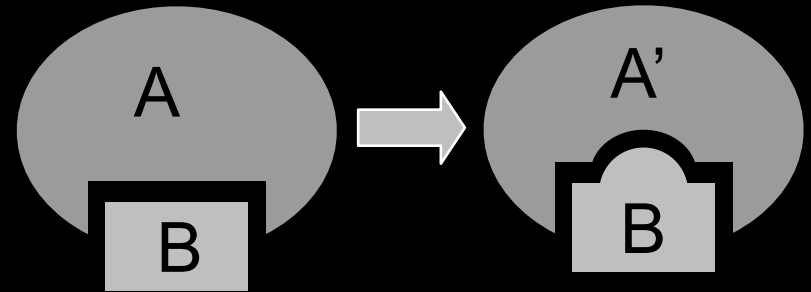
Tanja Kortemme
UCSF

Protein Interface Design - Concepts

Optimize Interaction

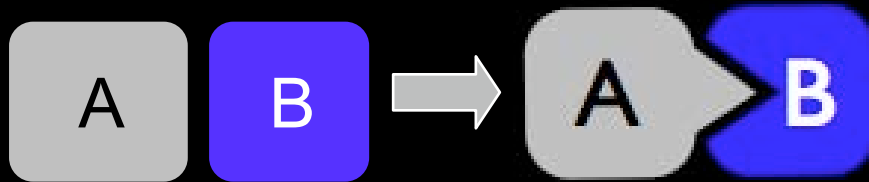


Altered Specificity



Protein Interface Design

- optimize an interaction

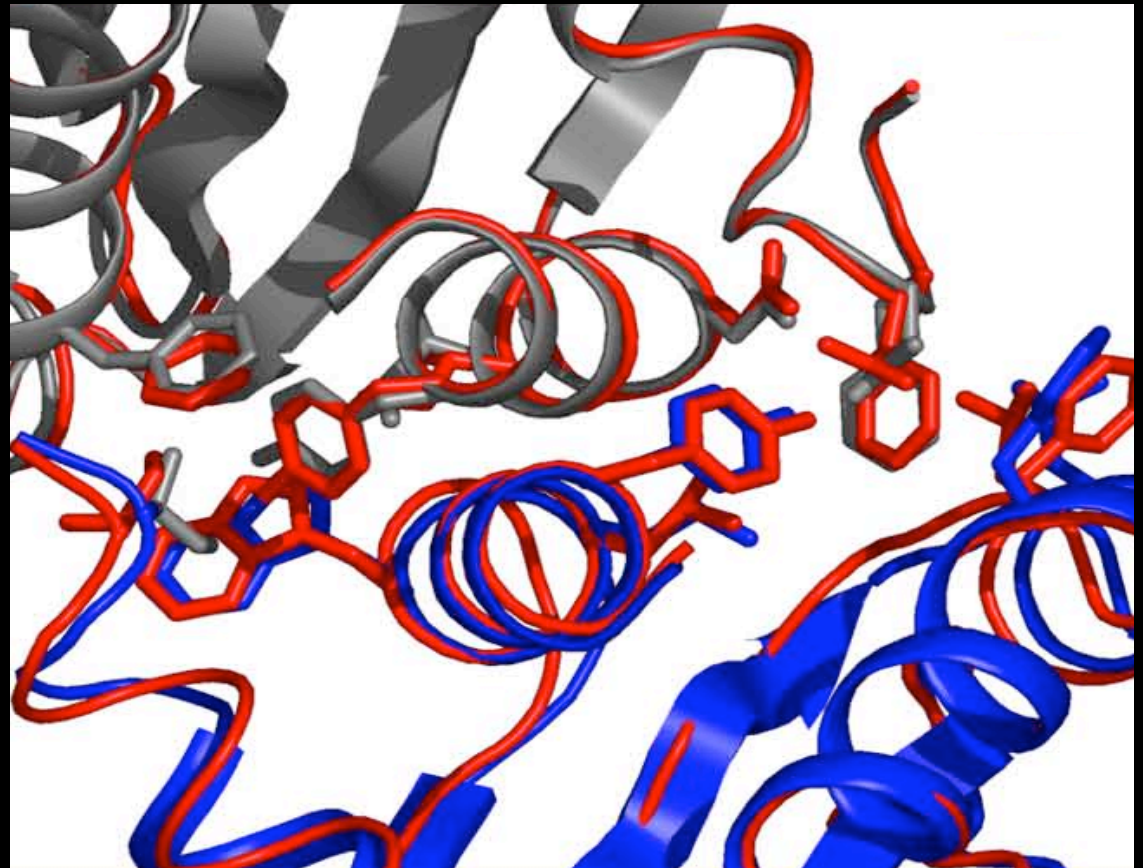


X-ray



DESIGN

8×10^{17} possible sequences
 6×10^{37} possible structures

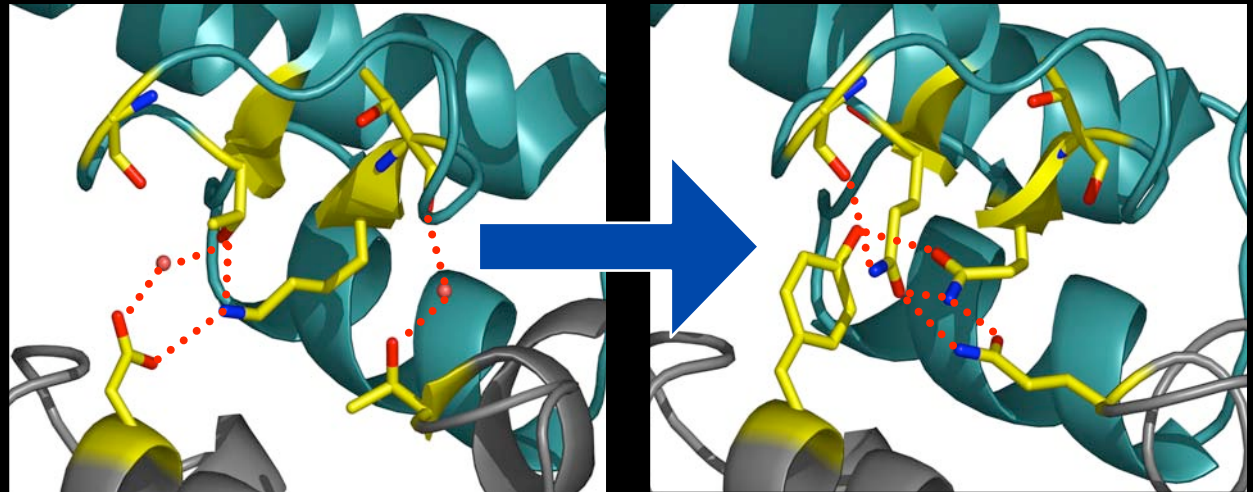
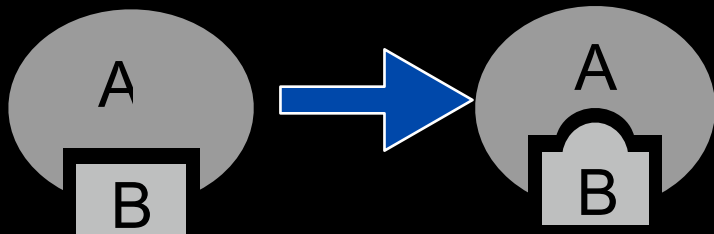
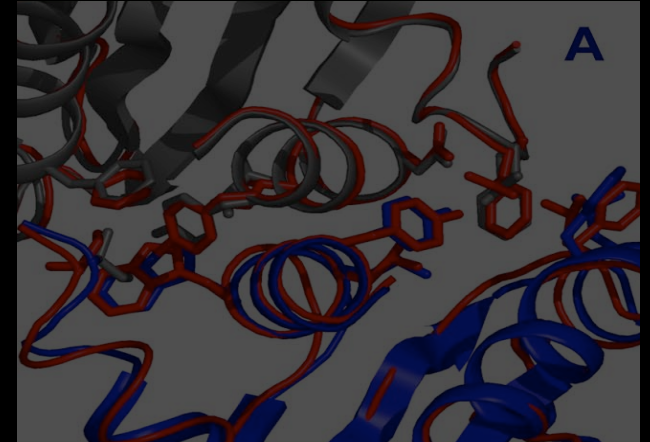


Mol Cell 2002

with Brett Chevalier & Barry Stoddard

Protein Interface Design

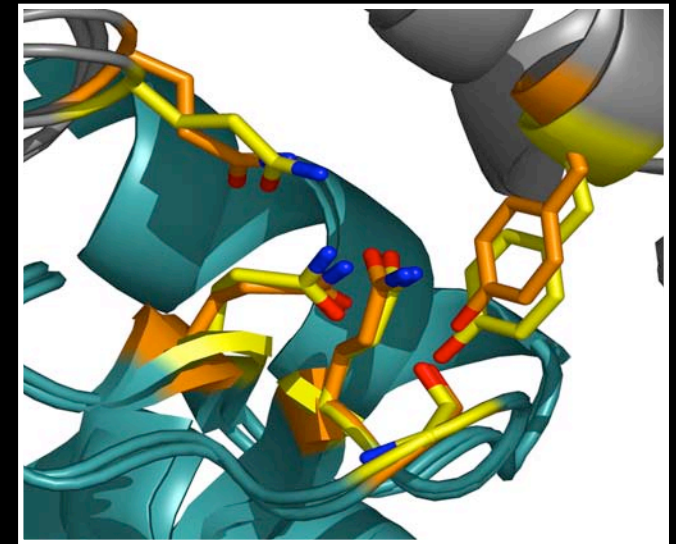
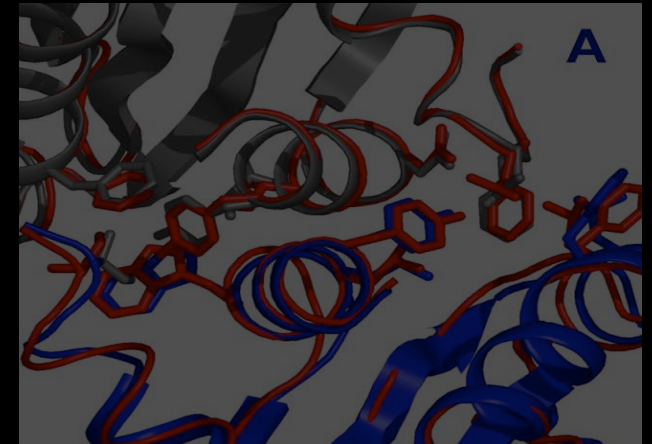
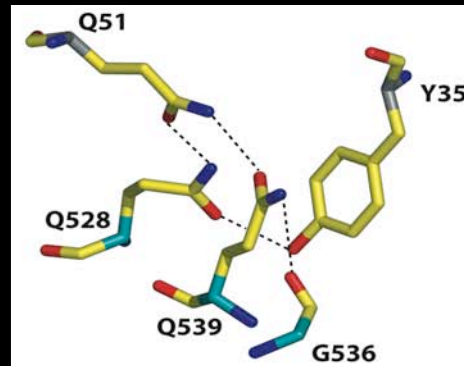
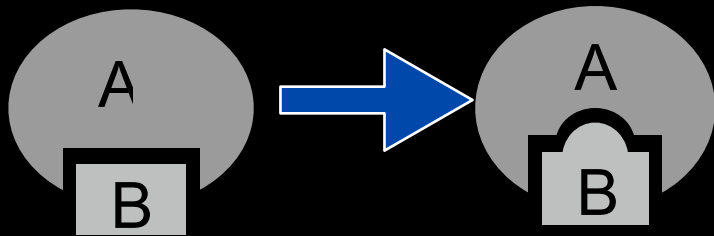
- optimize an interaction
- alter specificity:
remodel a hydrogen bonding network



*with Lukasz Joachimiak
J. Mol. Biol. 2006*

Protein Interface Design

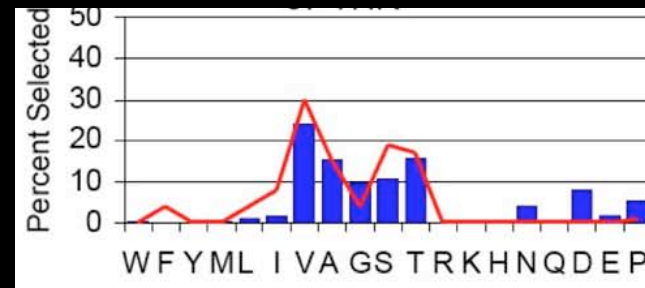
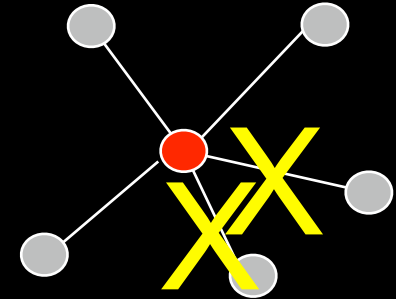
- optimize an interaction
- alter specificity:
remodel a hydrogen bonding network



with *Lukasz Joachimiak*
J. Mol. Biol. 2006

OUTLINE

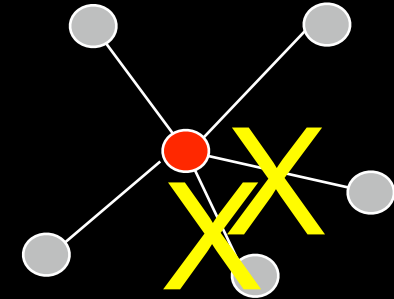
- reengineer protein interaction specificity
- flexible backbone design to predict tolerated sequences & libraries



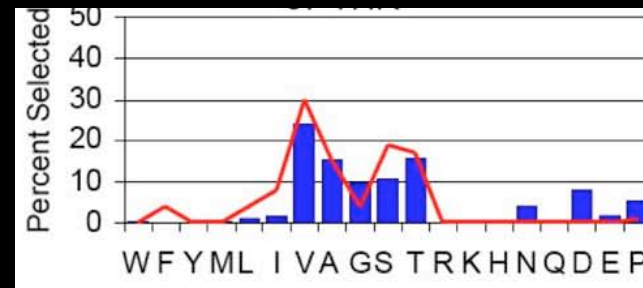
Elisabeth Humphris & Colin Smith

OUTLINE

- reengineer protein interaction specificity

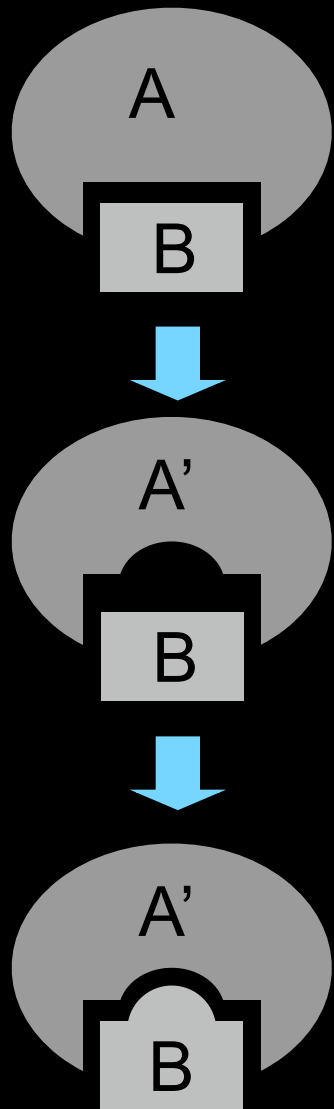


- flexible backbone design to predict tolerated sequences & libraries



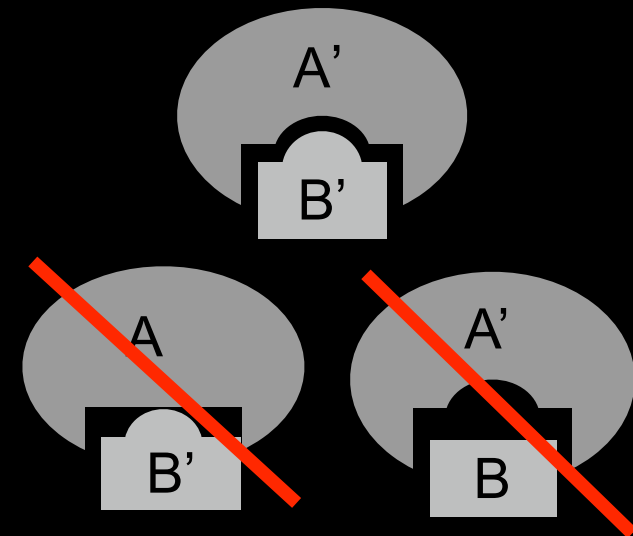
Elisabeth Humphris & Colin Smith

Computational Second Site Suppressor strategy to REDESIGN PROTEIN INTERACTION



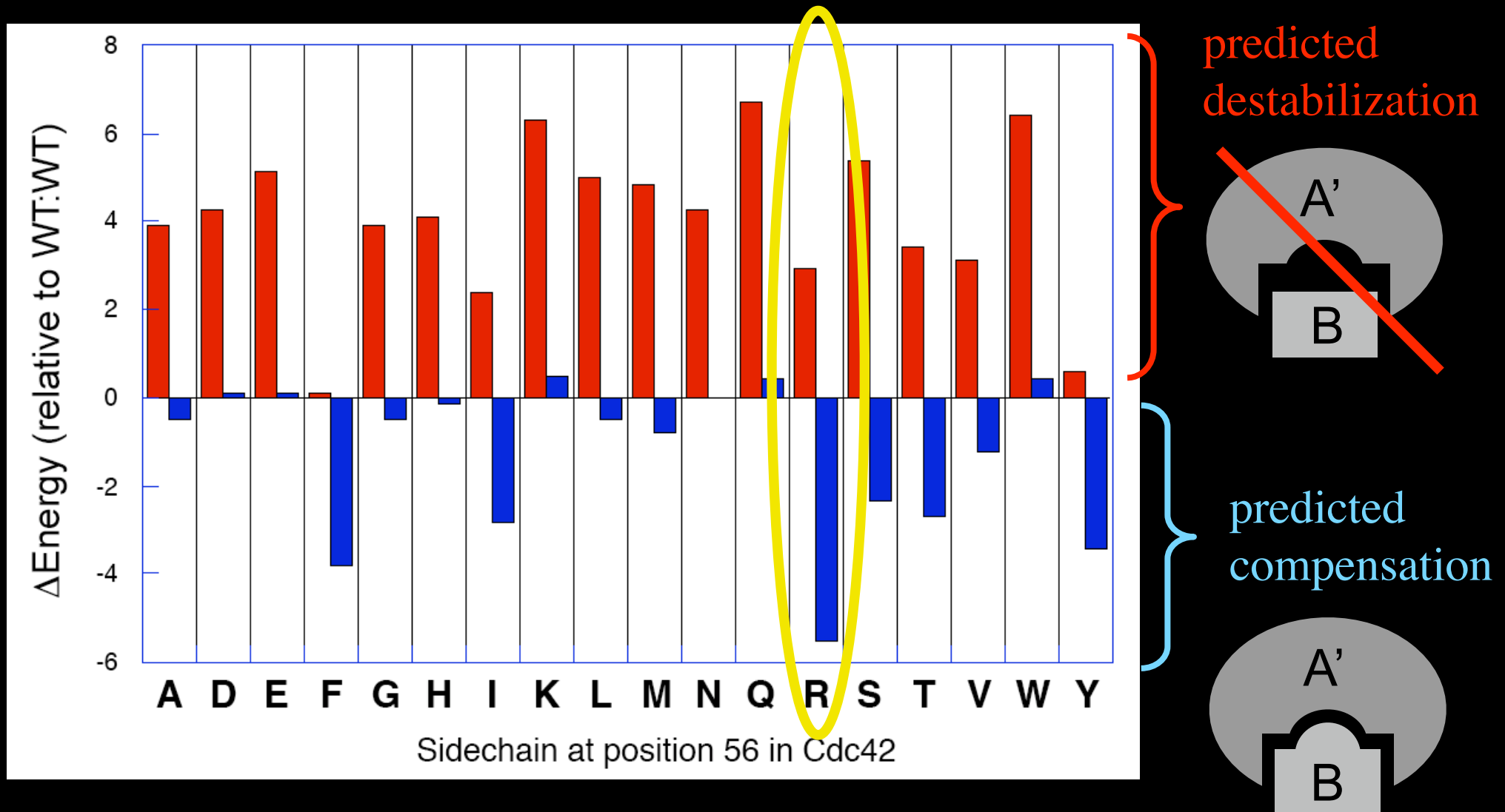
- computationally screen for destabilizing mutations in partner A
- redesign interface on partner B to compensate

select for:



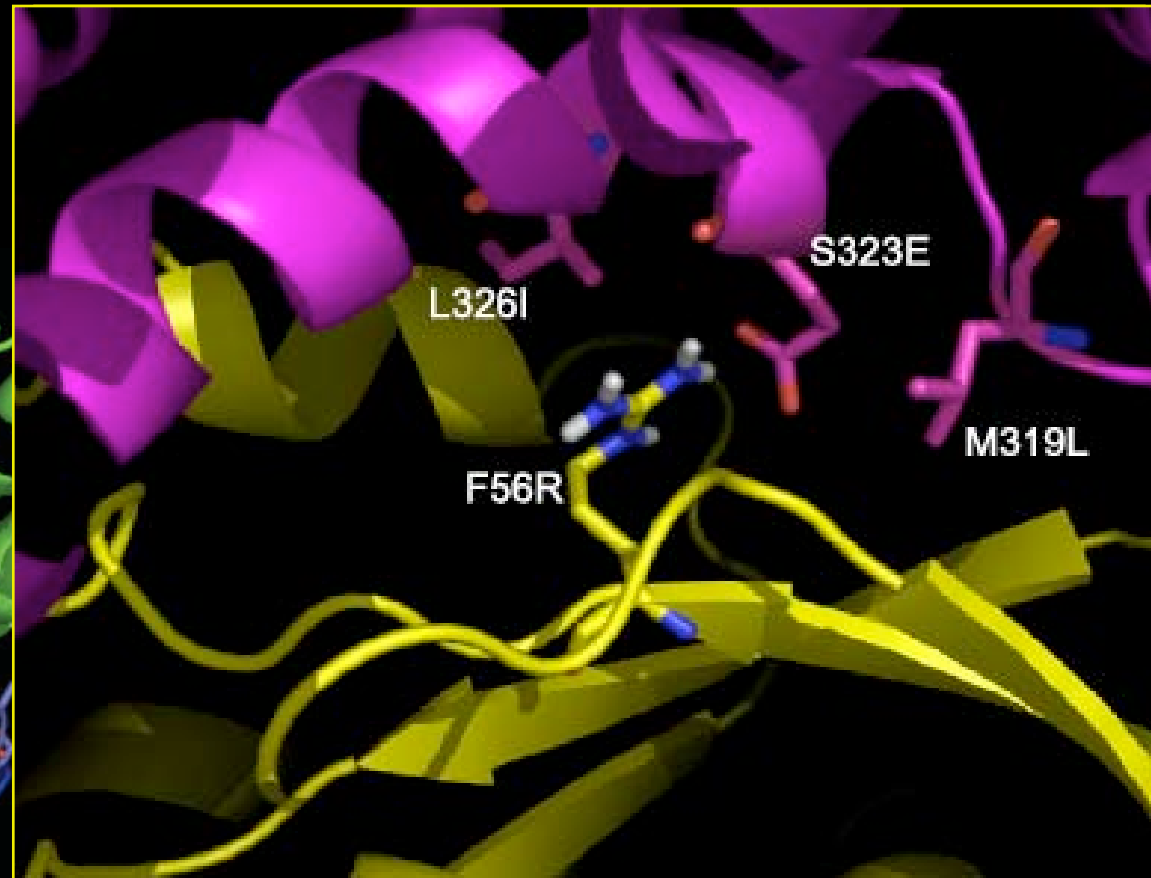
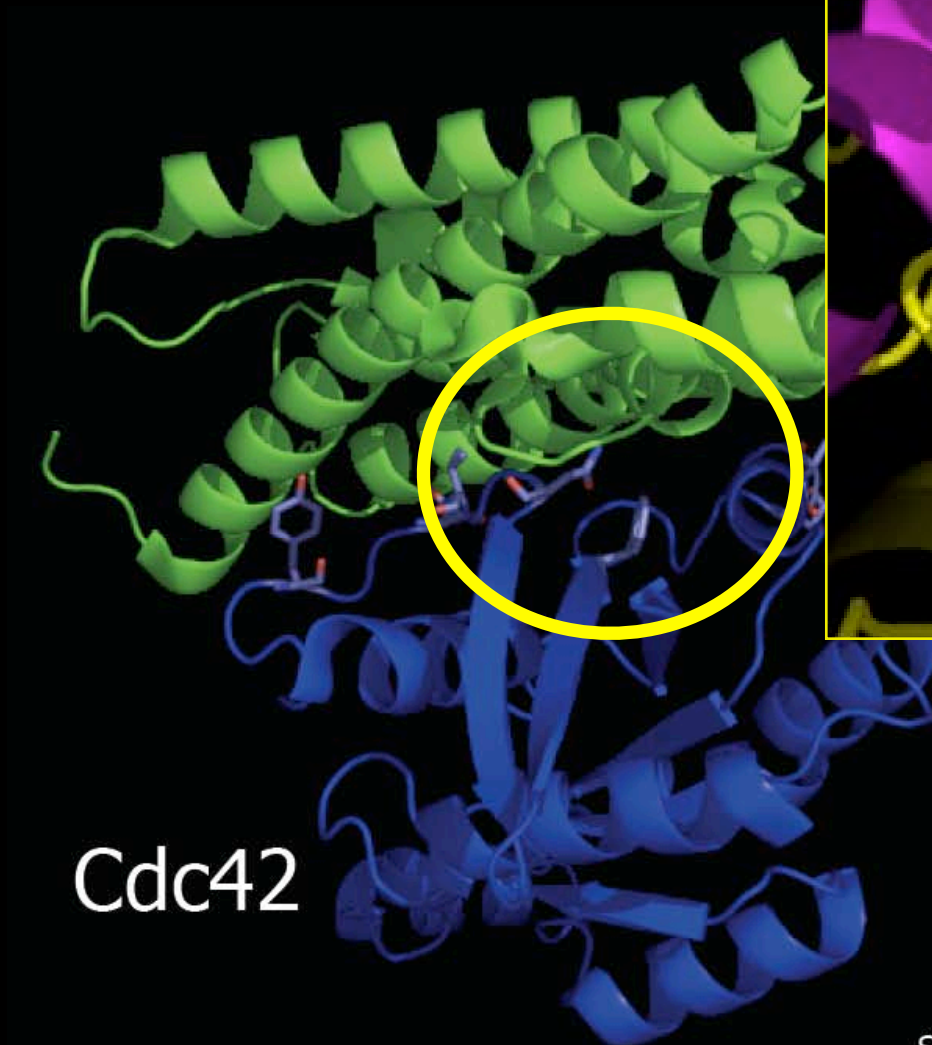
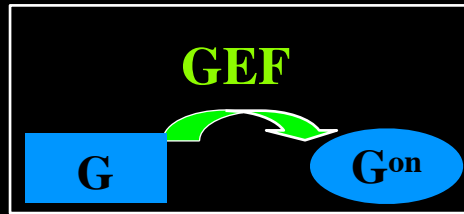
*Kortemme, Joachimiak et al.
Nature Struct. Mol. Biol. 2004*

Computational Second Site Suppressor Strategy



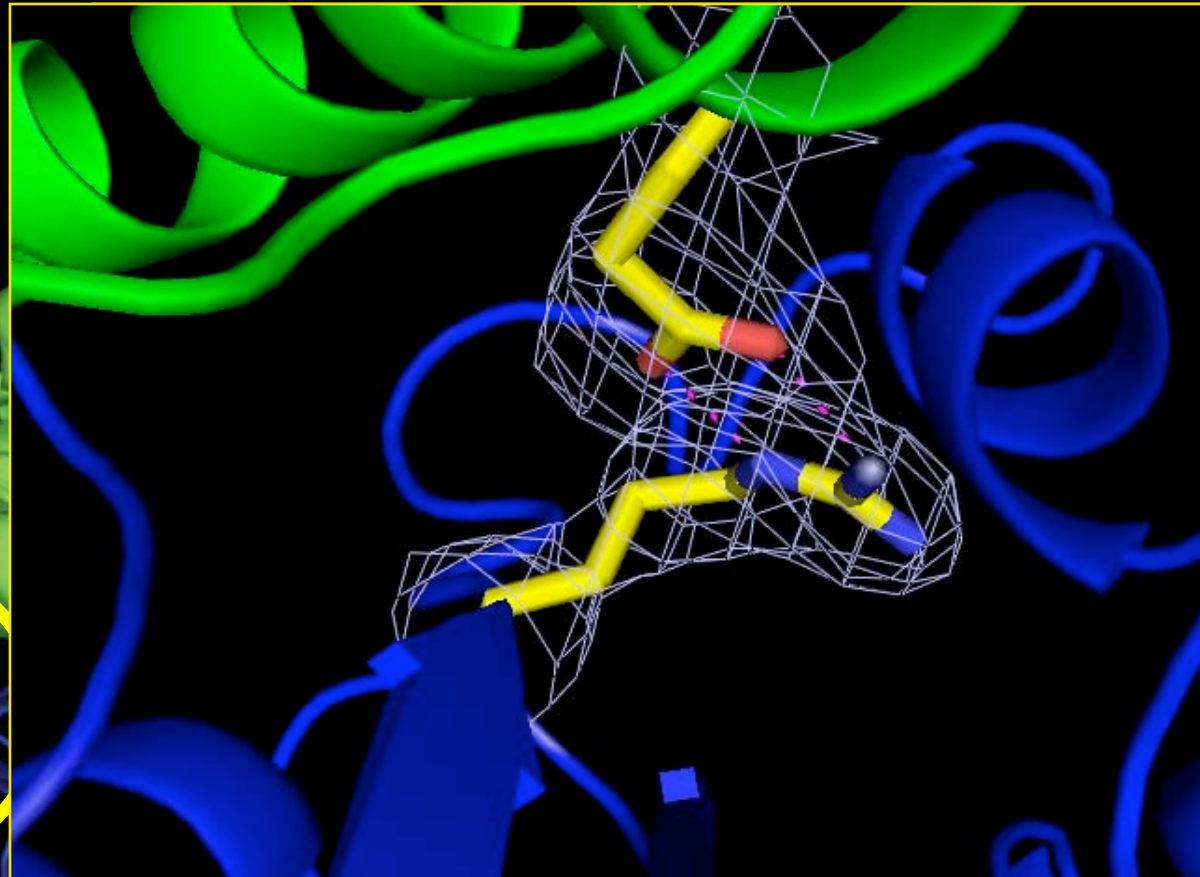
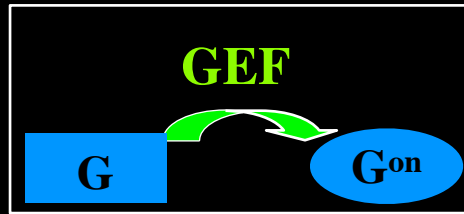
Greg Kapp

Design of new GTPase/GEF pairs



(predicted)

Design of new GTPase/GEF pairs



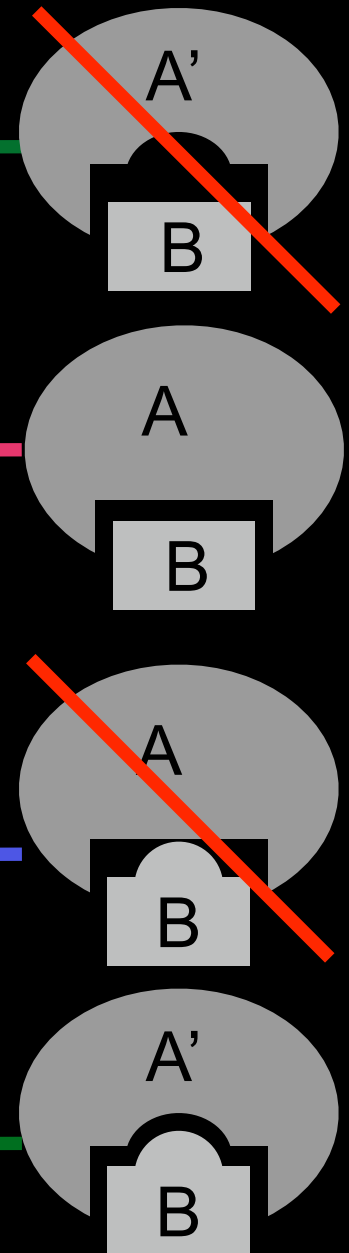
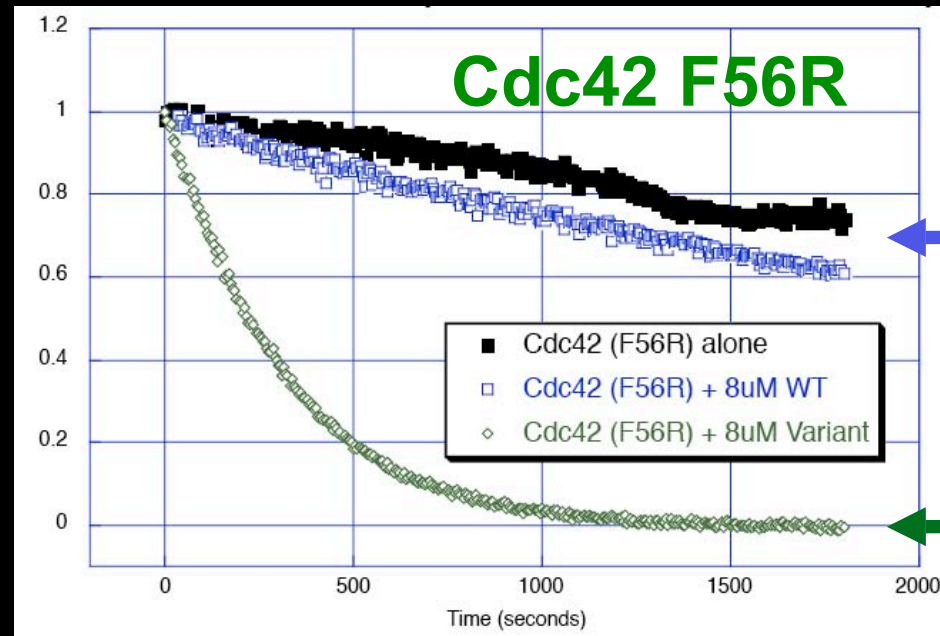
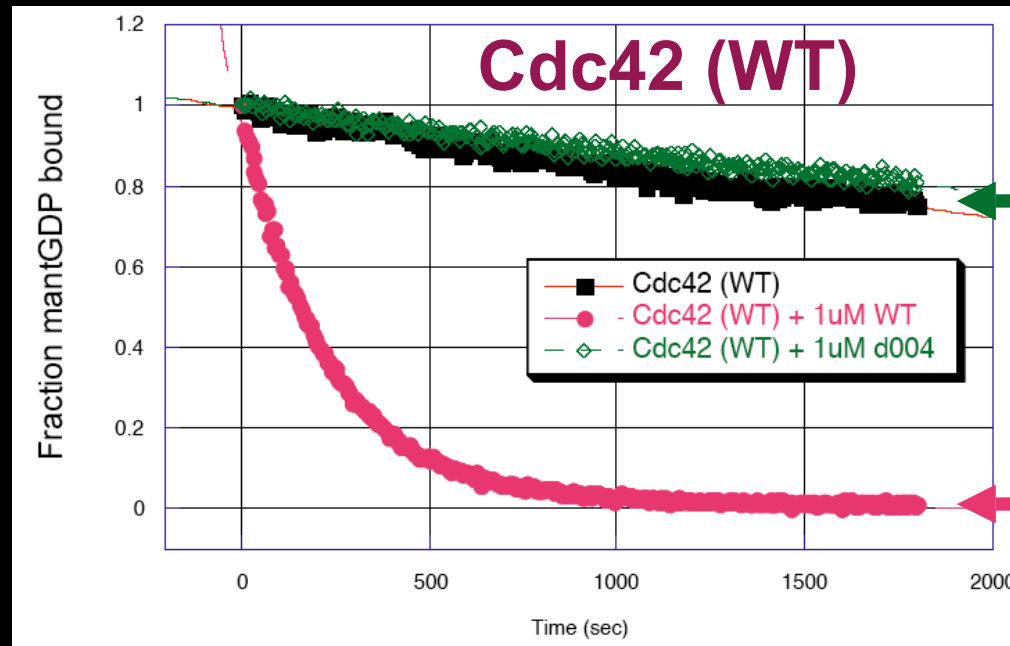
(X-ray)

In vitro activity of a designed GTPase/GEF pair

Uncatalyzed
(black)
and GEF catalyzed
(colours)
dissociation of
mantGDP
monitored by
fluorescence

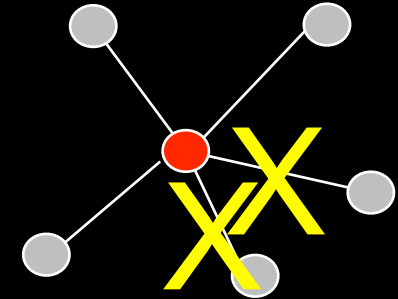


Greg Kapp

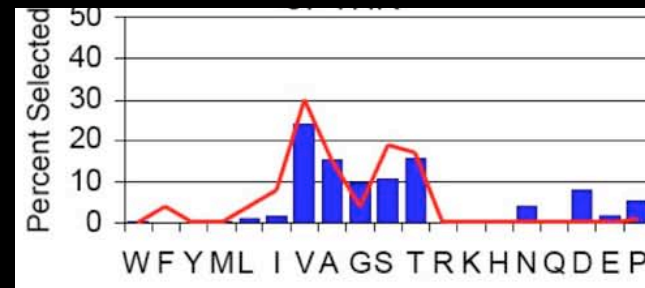


OUTLINE

- reengineer protein interaction specificity



- flexible backbone design to predict tolerated sequences & libraries

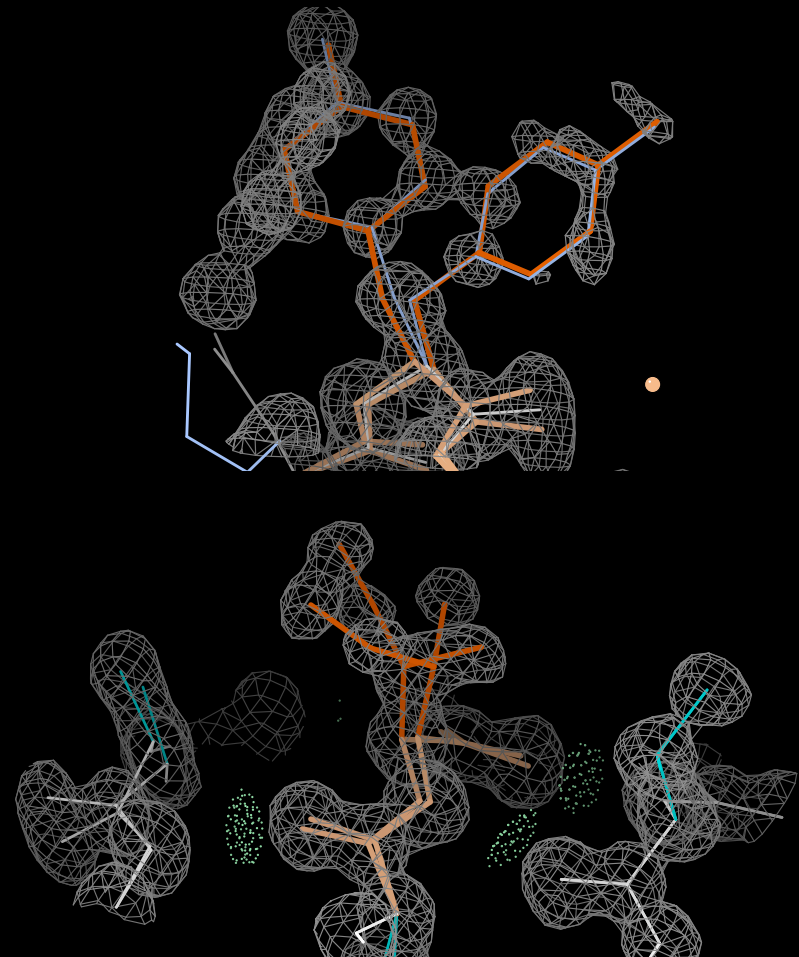
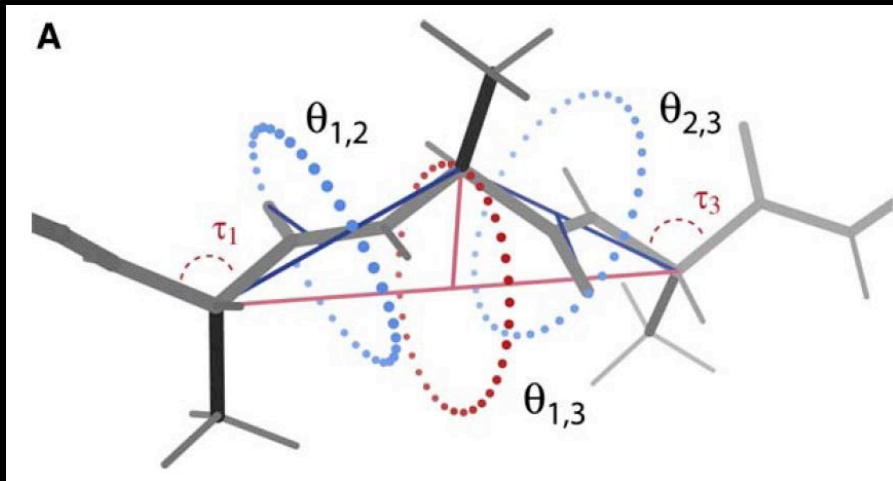


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- *proteins can undergo conformational changes in response to mutation or binding their partners*
 - what are good models to capture conformational variability?
 - how do we test these?

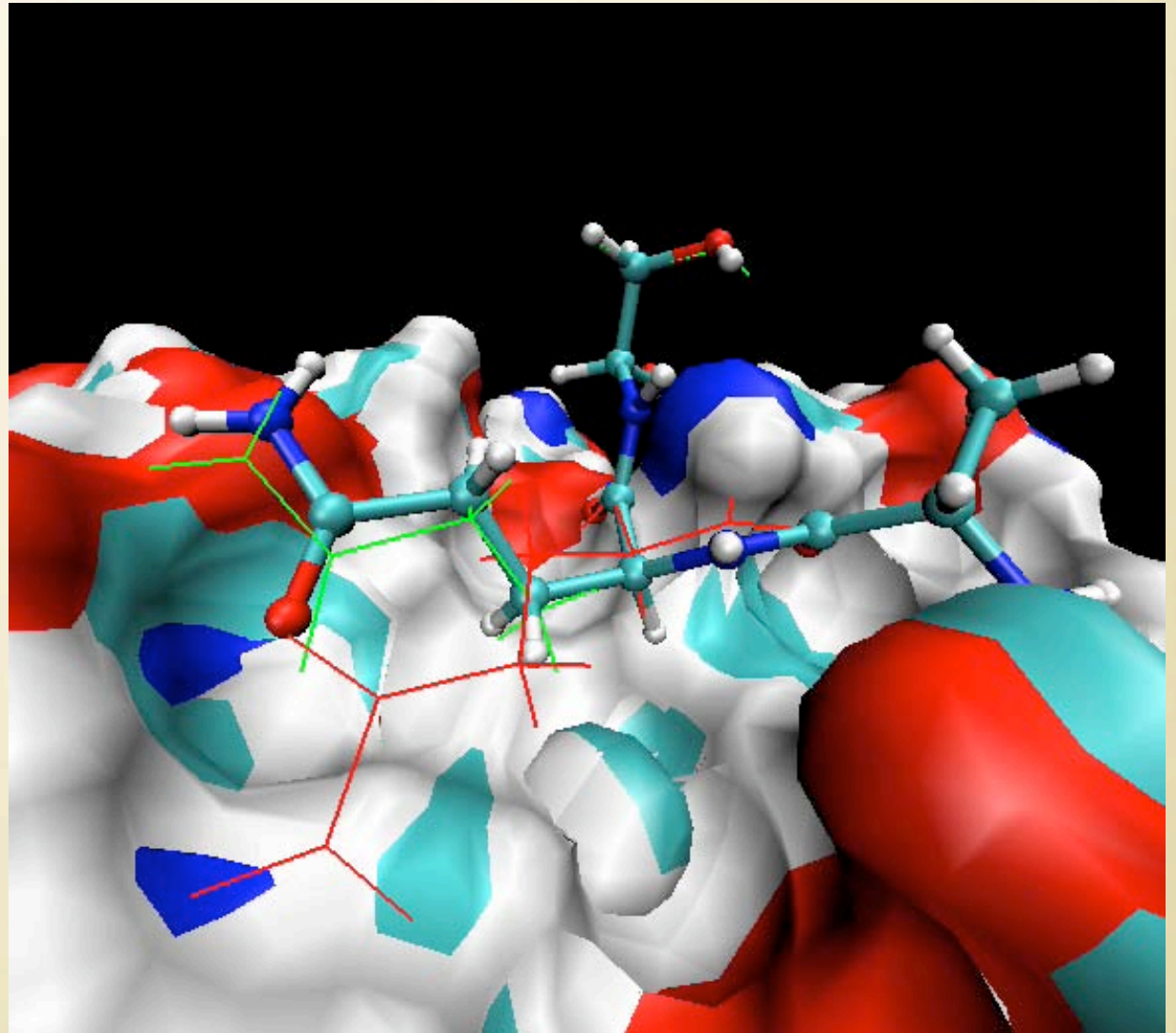
Modeling backbone flexibility at high resolution

motivated by a paper by the Richardson lab/Ian Davis:
“backrub” motion as a common type of local plasticity in the
backbone (Davis *et al.*, Structure 2006)



BACKRUB SIMULATION IN ROSETTA

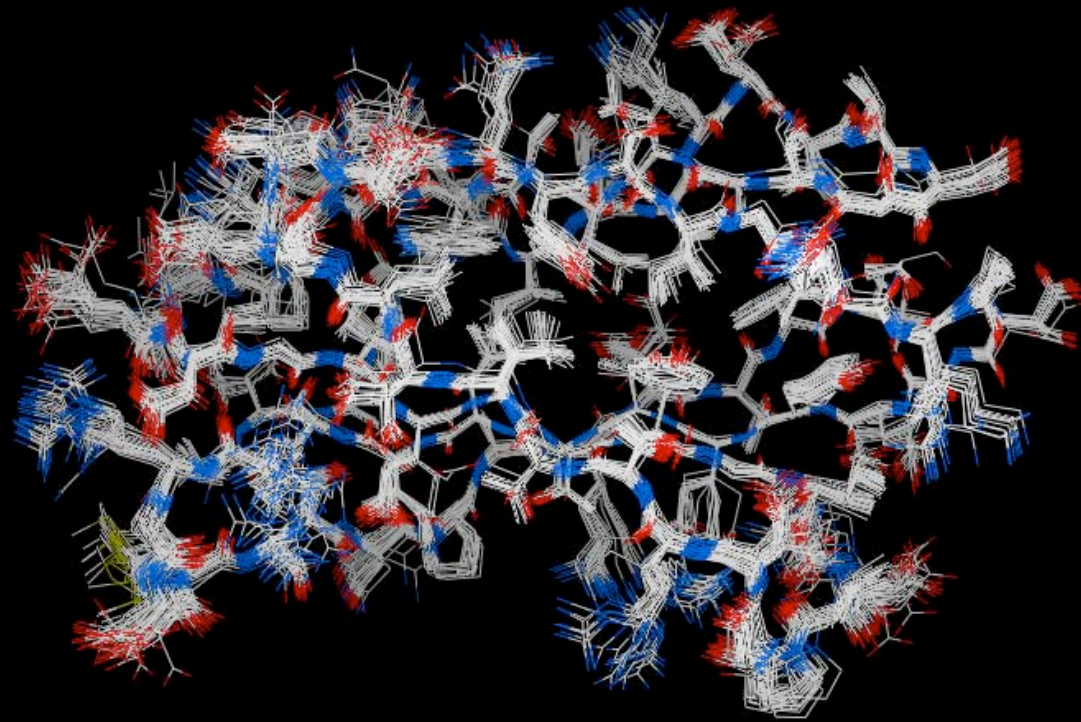
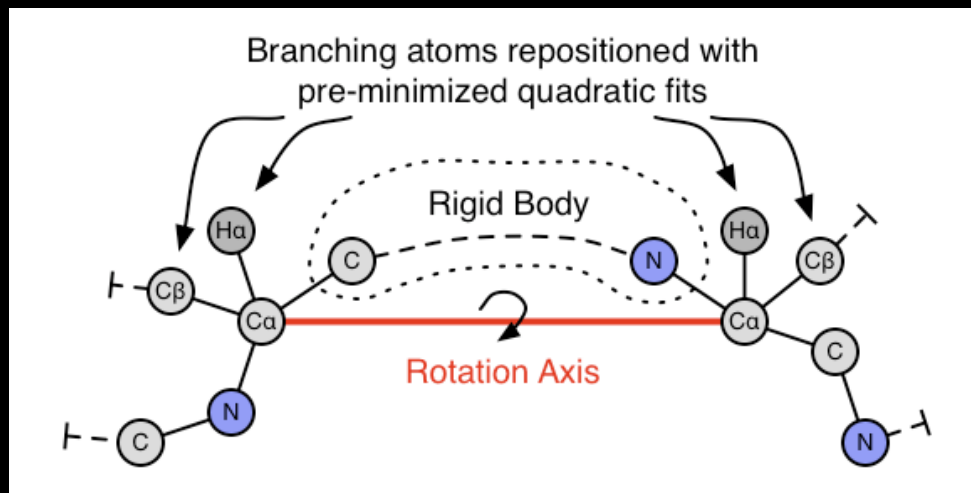
- coupled backbone/
rotamer motions



Colin Smith

Modeling backbone flexibility at high resolution

generalized backrub ($i, i+2$ up to $i, i+11$) as ROSETTA
Monte-Carlo move
(also for loop modeling)

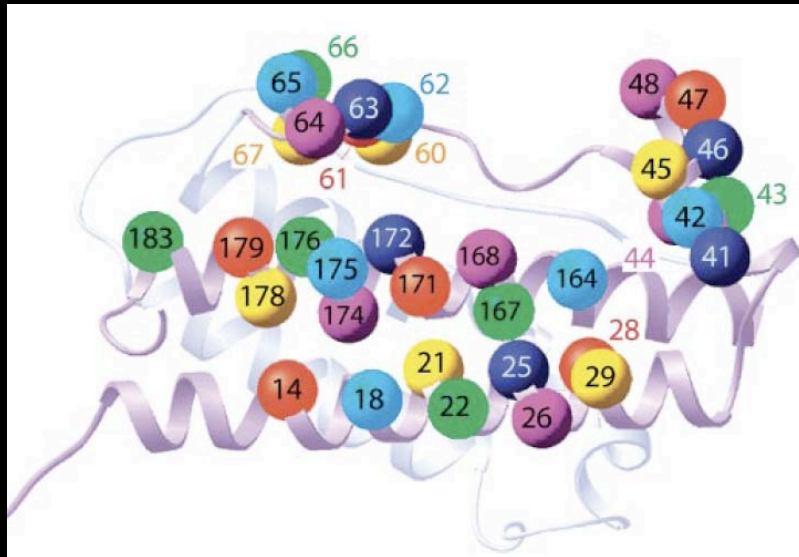


Colin Smith

Comprehensive and Quantitative Mapping of Energy Landscapes for Protein-Protein Interactions by Rapid Combinatorial Scanning*^S†

Received for publication, April 20, 2006, and in revised form, June 6, 2006 Published, JBC Papers in Press, June 8, 2006, DOI 10.1074/jbc.M603826200

Gábor Pál^{‡1}, Jean-Louis K. Kouadio[‡], Dean R. Artis⁵, Anthony A. Kossiakoff^{‡2}, and Sachdev S. Sidhu⁵³



selection from comprehensive libraries
for sets of 5-6 residues each

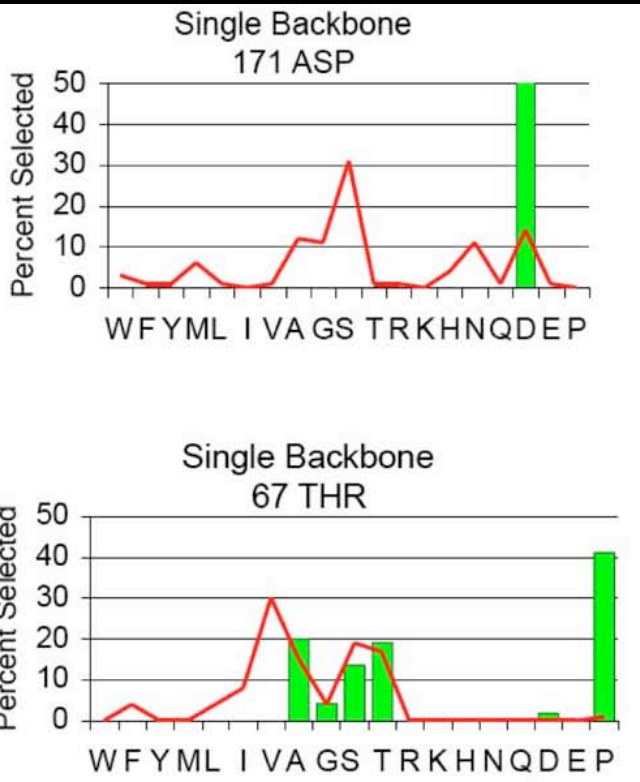
- for folding
- for receptor binding

major results:

- many substitutions tolerated
- patterns could not have been predicted from alignments

Predicting sets of tolerated sequences

HGH phage display results (*Pal et al., J. Biol. Chem. 2006*)



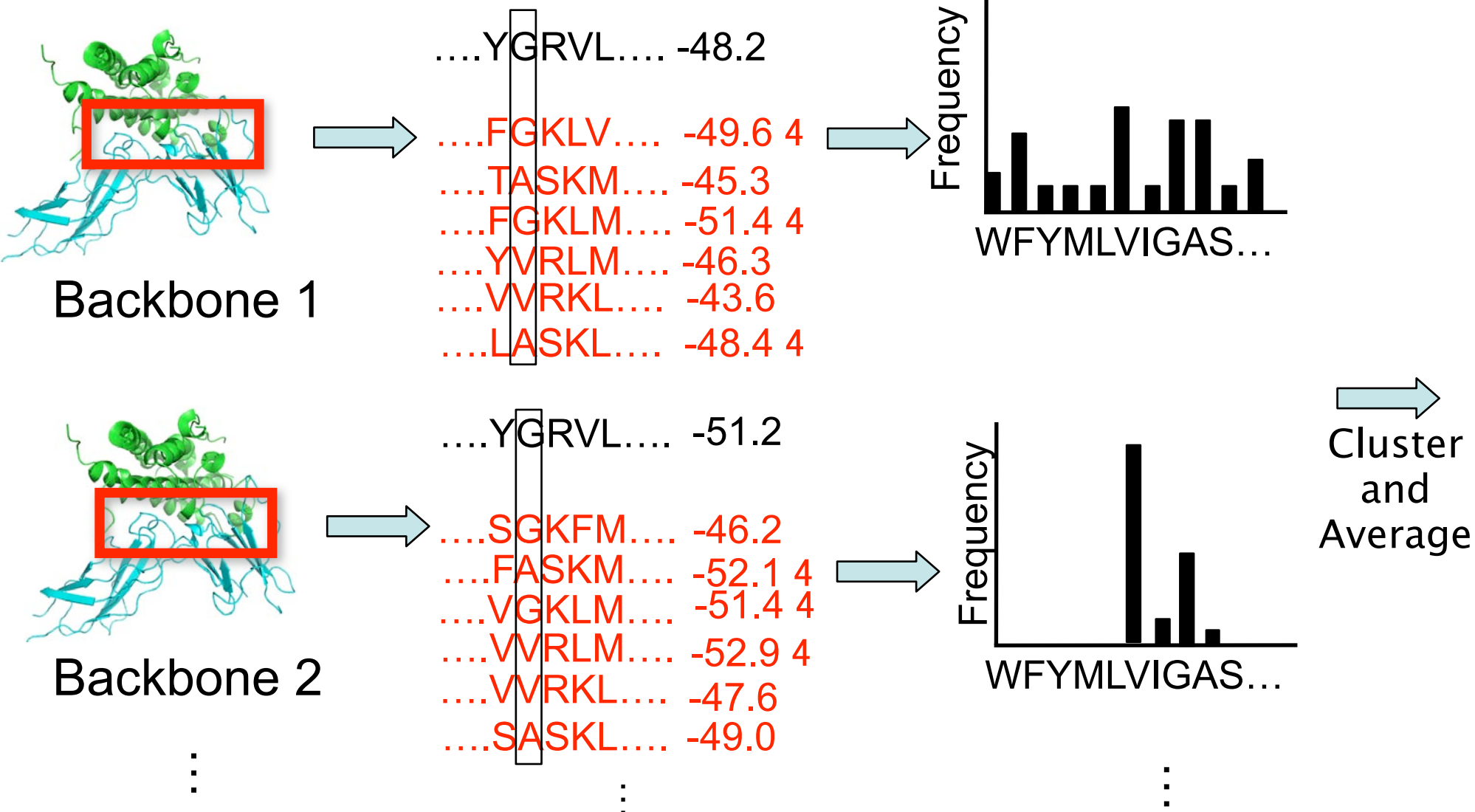
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Ensemble of 170 backrub-generated structure



Compile Profiles for Each Backbone

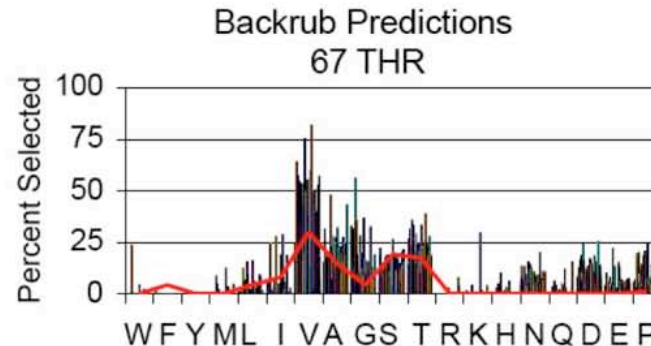
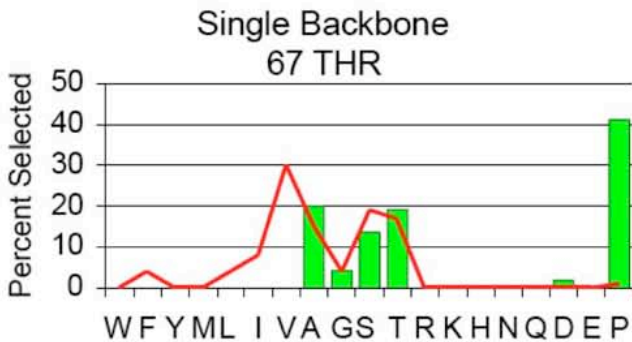
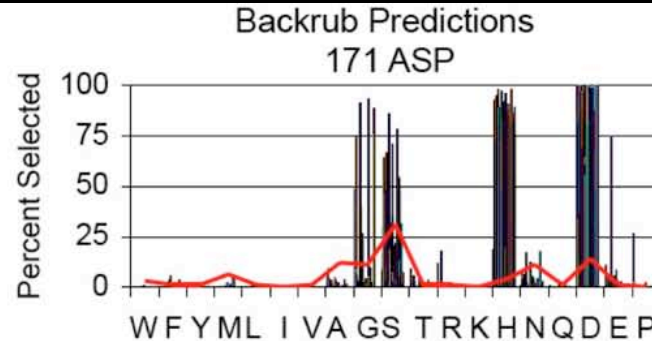
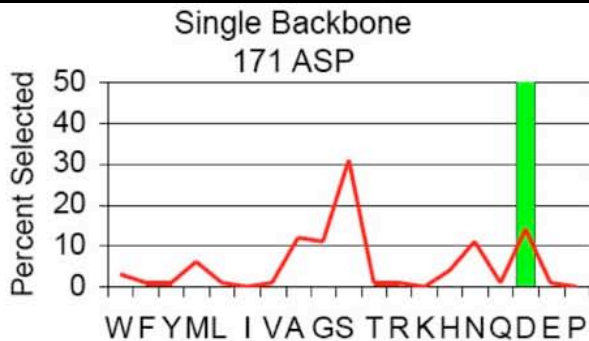
Everything Scoring “Better” Than Native



(Additional Threshold on Folding within 5% of wild type value)

Predicting sets of tolerated sequences

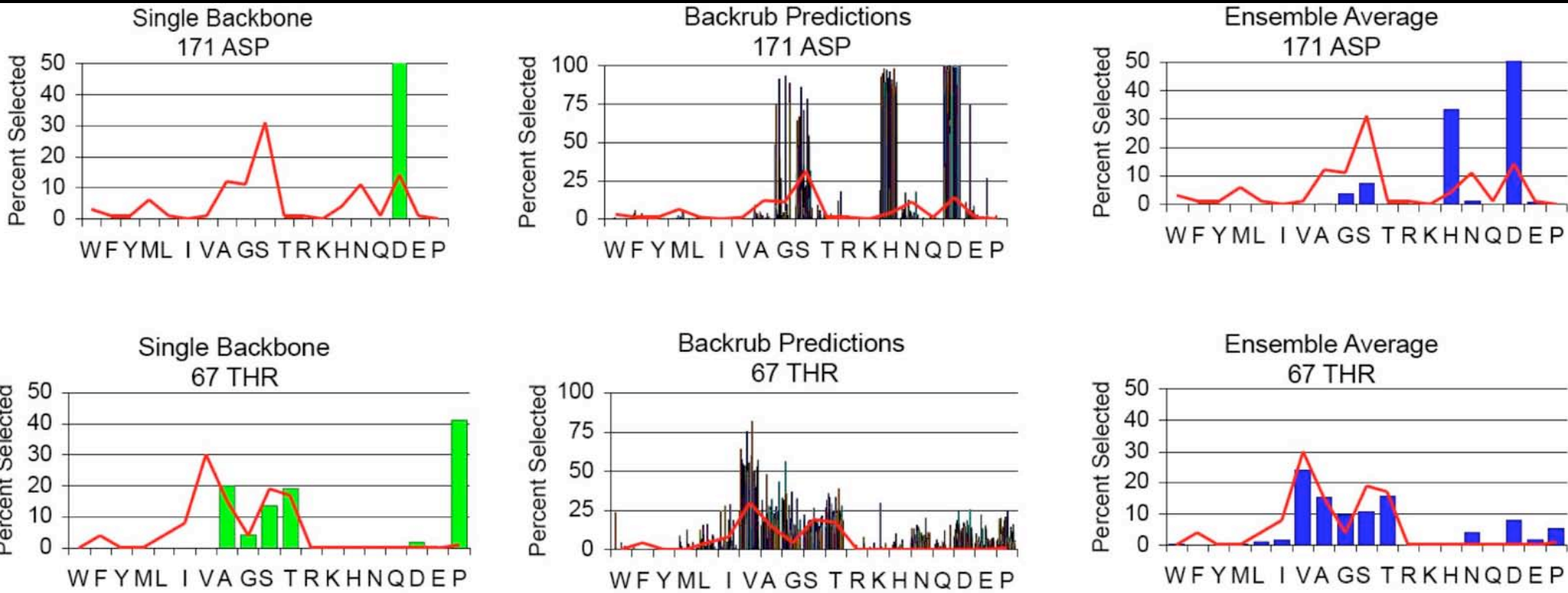
HGH phage display results (*Pal et al., J. Biol. Chem. 2006*)



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Predicting sets of tolerated sequences

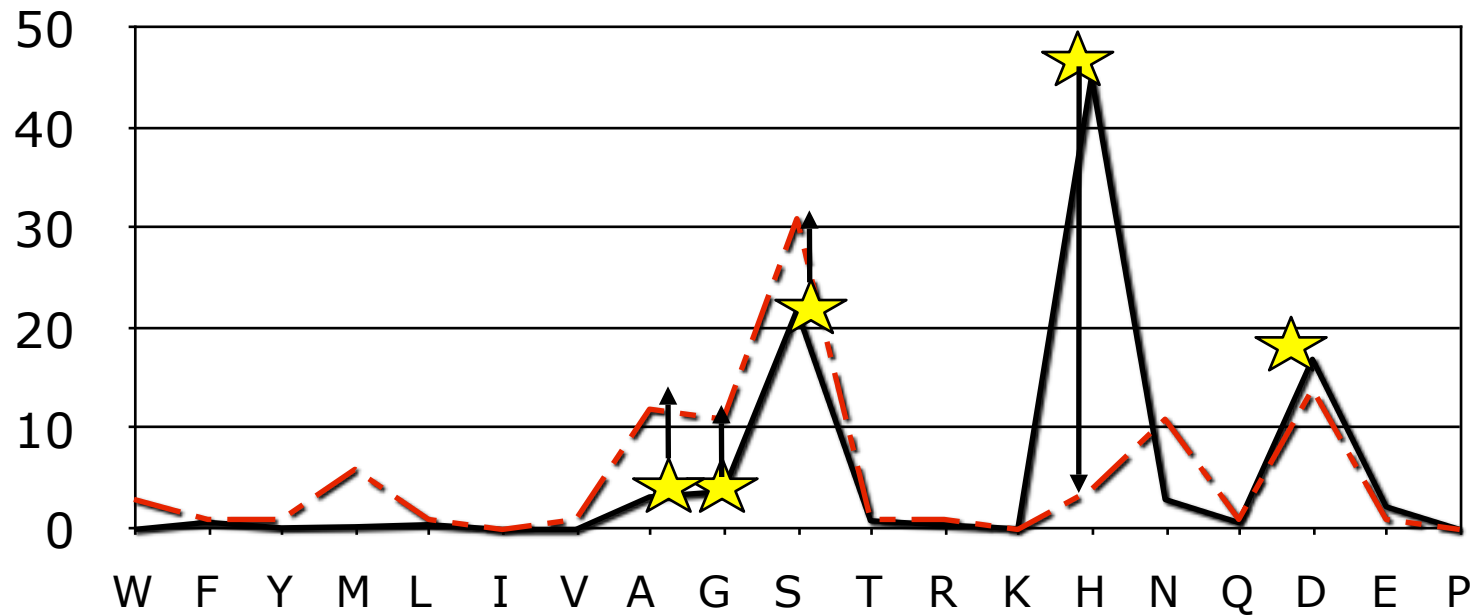
HGH phage display results (*Pal et al., J. Biol. Chem. 2006*)



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How well would computational predictions do for designing a library?

Pick 5 most frequent predicted amino acid types from each profile



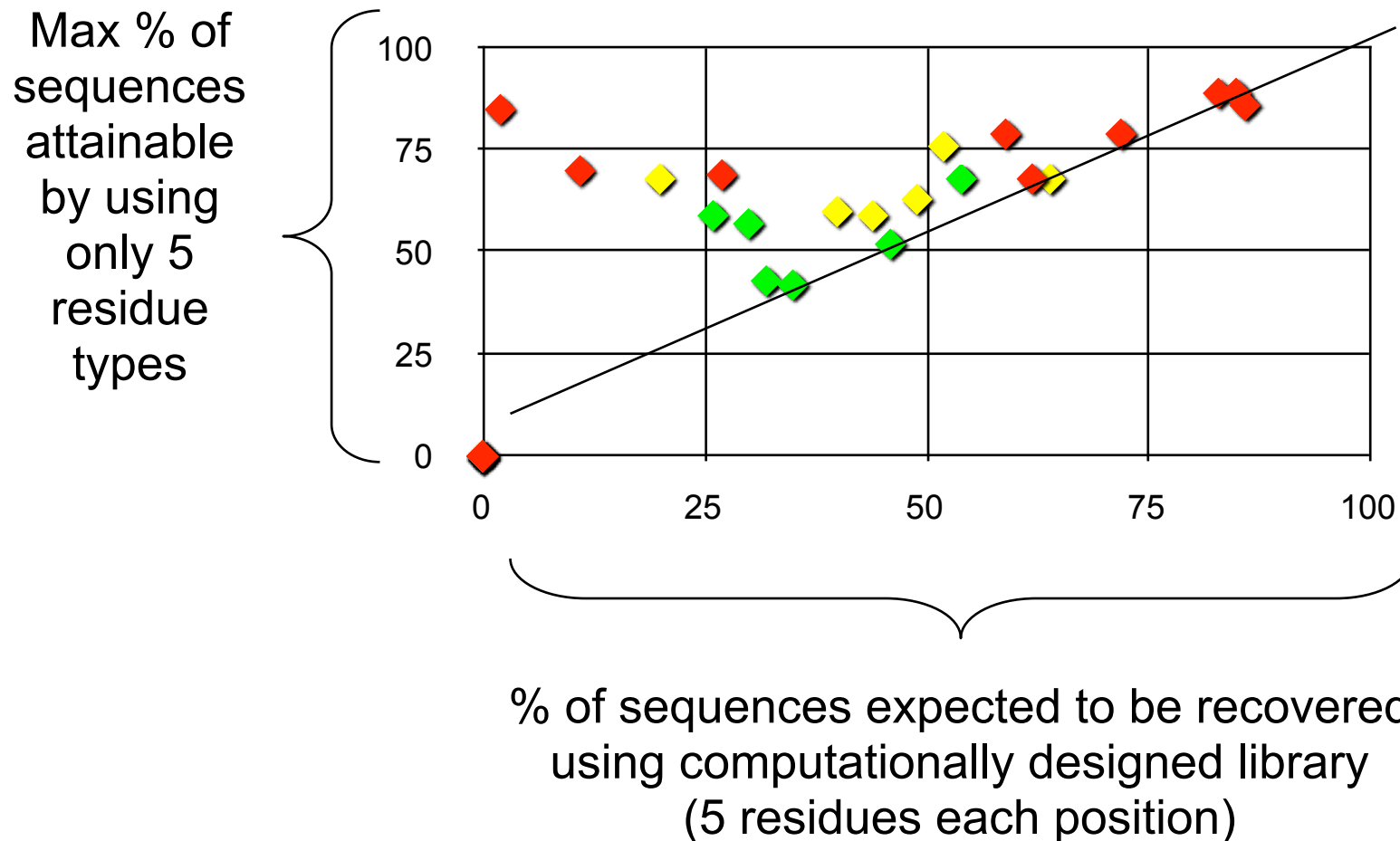
Computation: H, D, S, G, A

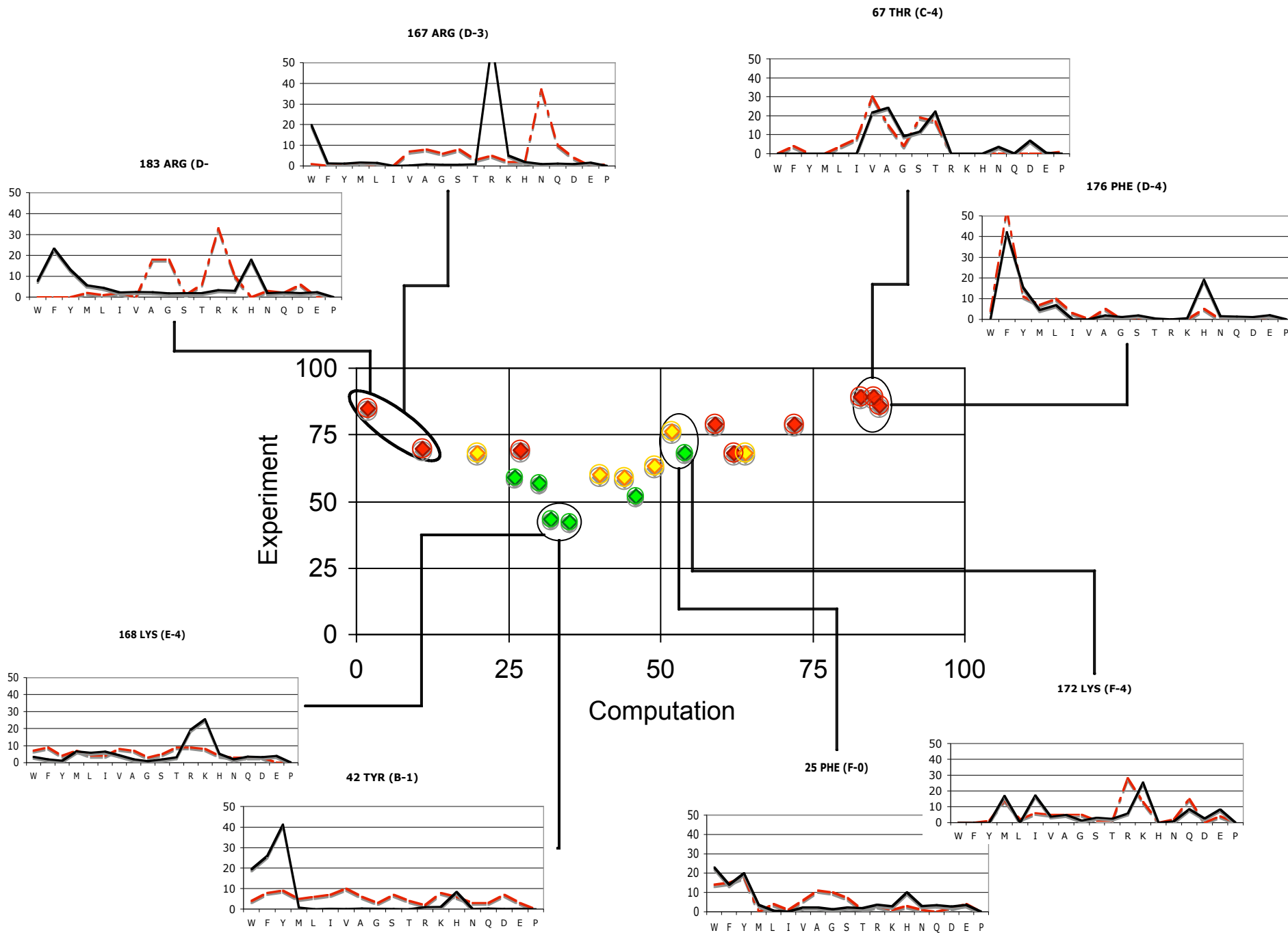
$$4 + 14 + 31 + 11 + 12 = 72 \quad \star$$

Max Seen: S, D, N, G, A

$$31 + 14 + 12 + 11 + 11 = 79 \quad \text{---}$$

Top 5 Residues Selected Computationally For Remaining 20 positions

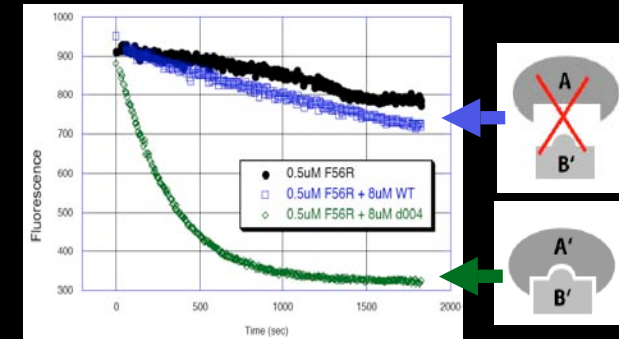




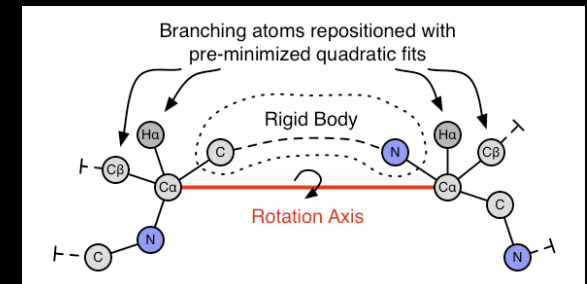
Conclusions

Reengineering sets of interactions

- GTPase/GEF pairs

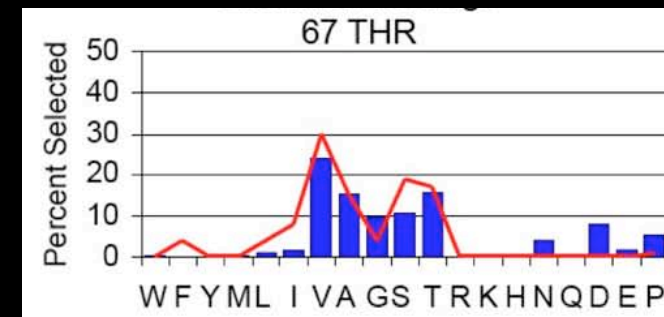


Backrub model for sampling local flexibility



Prediction of tolerated protein sequences using ensembles

- sequence plasticity & libraries



Thanks to:



Alfred P. Sloan Foundation, Sandler Family
NIH, NSF
Student Fellowships:
NSF, Genentech/Sandler, DOD, ARCS

Computational Models

- *Colin Smith
- *Greg Friedland
- *Dan Mandell
- *Anthony Linares
- *Mariana Babor
- *Ryan Ritterson

Sequence Plasticity

- *Elisabeth Humphris

Sets of Interactions

Greg Kapp
Cristina Melero
Brian Yeh, Attila Remenyi,
Wendell Lim

Protein Evolution

Matt Eames

Networks

Rich Oberdorf